REVIEW



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Aging, immunosenescence and membrane rafts: the lipid connection

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Abstract

The decreased efficiency of immune responses in older people is partly a consequence of alterations in T lymphocyte functions caused by modifications in the early events of signal transduction. Several alterations in the signaling pathways of T lymphocytes have been described in older humans and animals. A unifying cause could be modifications in the physicochemical properties of the plasma membrane resulting from changes in its lipid composition and the distribution and function of lipid rafts (LR). The latter serve to assemble the initial components of the signaling cascade. Accumulating data suggest that the function of plasma membrane LR is altered with aging; we hypothesize that this would significantly contribute to immune dysregulation. The role of aging and cholesterol in LR functions in T lymphocytes is reviewed and discussed here.

Keywords: Cholesterol, Immune synapse, Immunosenescence, Lipid rafts, Signal transduction, T cells

Review

Aging is a complex and multifactorial process. One of the outcomes of aging is the appearance of numerous diseases that are caused by alterations in different physiological systems. These changes appear to occur progressively throughout life and become more pronounced with age. Alterations in the homeostatic status of the body with aging decrease its capacity to cope with daily challenges, resulting in an increased, overall level of physiological stress and a greater demand on bodily reserves. The immune system is particularly affected by the process of aging [1], especially regarding the numbers and functions of T lymphocytes. Studies of older humans and animals have revealed that the most noticeably altered T cell functions are the production of IL-2 and the clonal expansion of effector and memory T cells [2]. These functions are the outcome of the activation of T cells and it can be hypothesized that defects or alterations in the proximal events of T cell activation will strongly affect the efficiency of the immune response. In

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What is Immunosenescence?

As life expectancy of the populations of industrialized nations continues to increase, the emergence of many diseases associated with aging has become a major medical and economic concern. For instance, the incidence of infections, cancers and chronic inflammatory diseases such as atherosclerosis and neurodegenerative diseases



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which increase with age has become an important human and cost-related problem [9]. Although the underlying causes relating aging to increased pathologies is not fully understood, it is recognized that alterations of the immune system play an important role [10]. The immune system is a complex interactive system composed of many different cells and cell subpopulations that are altered with age, but not all to the same extent, and thus do not contribute equally to immunosenescence. This should be conceptualized as dysregulation of a system constantly trying to adapt and to maintain homeostasis in the face of inputs and outputs which are still only crudely defined [11]. Although numerous studies on age-related immune alterations have been published, the conclusions remain controversial due to differences including those between species and the lack of a definition of physiological aging. Furthermore, the exclusion of some latent disease states, nutritional, genetic and environmental differences add to the complexity of understanding the mechanisms of the physiological changes and their overall contributions to aging [12]. However, the direct clinical consequences of decreased immune responses with aging seem quite clear. These are revealed by the increased incidence and severity of infections, cancers and autoimmune disorders [10,13] and can be related to the hallmark of immunosenescence which is altered T cell function with aging [14]. Additional changes in immune system components have been described, but they are much less marked and may often be secondary to changes in T cells. The changes include not only the T cell-dependent B cell responses, but also innate components sensitive to T cell feedback, especially APC [15]. Although age-related changes in the immune response are multifactorial, it is reasonable to assume that an alteration in T lymphocyte activation is a central issue in age-related modifications of the immune response.

Changes in T lymphocyte activation with aging *T cell signaling via T cell receptor and CD28*

T cells are activated by antigen presented by APC [16]. The most efficient APC are dendritic cells, although monocytes/macrophages and B cells are also able to present antigens. APC presentation within the context of major histocompatibility restriction triggers a cascade of signaling events in naive T cells that culminates in the activation of a set of genes resulting in the synthesis and secretion of IL-2 and many other cytokines and factors (Figure 1). In addition, activated T cells initiate immune responses and traverse differentiation pathways resulting in the development of different T cell subsets, according to the priming conditions [17] regulated by the cytokines and chemokines secreted in the immediate vicinity [7]. A universal requirement for any T cell response is marked proliferation to accomplish clonal expansion that is critical for generating T cells in sufficient numbers to eliminate the antigen. Memory T cells possess different stimulatory requirements than naïve T cells, but the efficiency of both naïve and memory cell responses is diminished with age.

The primary receptor for T cell activation is the T cell receptor (TCR) which delivers "signal 1". However, this is not sufficient for full T cell activation and, to avoid anergy, T cells must receive a second signal delivered by a plethora of co-receptors [17]. A number of these have been described [16] but, functionally, the most important of these seems to be CD28 which, by recognizing CD80 and CD86 ligands on APC, delivers "signal 2" (Figure 1) that is essential to sustain T cell activation [18]. In addition to its activity as a co-stimulatory molecule, CD28 is involved in re-expression of the TCR, recruitment and stabilization of T cell LR to the immune synapse and the activation of plasma membrane lipid metabolism through upregulation of the PI-3 K signaling pathway [19,20]. Furthermore, CD28 co-stimulation synergizes with TCR activation and induces IL-2, IL-4, IL-5, TNF and granulocyte-macrophage colony stimulating factor (GM-CSF) production via NF-kB activation [19,21]. More co-stimulation is required for the activation of naïve cells than for memory cells. It is now well established that CD28 expression is decreased with aging in CD8⁺ and CD4⁺ lymphocytes with a bias towards a greater effect on CD8⁺ T cells [22].

Age-related alterations in T cell signaling

There are several age-associated alterations in these T cell activation pathways, as observed in experimental animals [23,24] and humans [2]. The most important changes occur in CD4⁺ T cells resulting in the decreased production of IL-2 and clonal expansion [3]. Although there are no changes in TCR number at the cell surface, the number of CD28 co-stimulatory molecules decreases with aging [25]. Due to the essential role of CD28 to prevent T cell anergy, the decrease in number of this receptor could affect T cell responses in aged humans. Nearly all the activity of the signaling pathways associated with TCR activation or IL-2 receptor response have been found to be altered with aging [26]. There is an alteration in the early steps of T cell activation including protein tyrosine phosphorylation, calcium mobilization and the translocation of protein kinase C to the plasma membrane. In addition, subsequent steps of the signaling pathways including the Raf-Ras-MAP kinase pathway are impaired. The decline in proximal and intermediate events of transmembrane signaling leads to the decreased activity of transcription factors, especially NF-kB and NF-AT [27] and results in altered production of cytokines with aging in T cells. This has been observed in the case of T helper-1-dependent cytokines such as IL-2, IL-6 and TNF- α production [28].



Lck regulation loop alterations with aging

Lately it became evident that one of the most important changes underlying altered TCR signaling is at the level of the Lck pathway (Figure 2A). Lck is at center stage of TCR signaling initiation [29]. Phosphorylation of the TCR by Lck initiates downstream signaling by creating binding sites that recruit the cytosolic kinase ZAP70 (Zeta-chain-associated protein kinase 70) to the cell membrane [30]. Lck is regulated by the C-terminal Src kinase (Csk) by transphosphorylation and by dephosphorylation by phosphatases. Lck cycles between an unphosphorylated (primed state), active and inactive states. The Lck C-terminal tyrosine Tyr505 is phosphorylated by Csk which allows the Lck C-terminus to bind its own SH2 domain, which closes the molecule. In contrast, the trans-autophosphorylation of Lck activation loop Tyr394 activates the kinase activity via rearrangements of the active site [31]. In resting cells equilibrium probably exists between the Lck phosphorylated on various Tyr sites resulting in various levels of kinase activity determined by the amount of Csk, CD45 and SHP-1 (Src homology 2 domain-containing protein tyrosinephosphatase 1) activities (Figure 2A). Reduced phosphatase activity within a close contact zone of Lck results in Lck activation by trans-autophosphorylation of Tyr394 [32]. However, it seems that some part of Lck is in an active form in resting T cells where a decrease of phosphatase activity will induce the signaling cascade as well as segregation in various membrane domains. Indeed, at basal states, Lck partitions preferentially into LR compartments where active and inactive Lck forms co-exist at different stoichiometry and interact with distinct pools of CD4 and Csk [33]. Thus, modulation of phosphatase activities in the close contact zone is an essential stage of TCR activation to release the already active Lck. Furthermore, in resting lymphocytes, Fyn has been shown to be the predominant Src-family tyrosine kinases (SFK) responsible for PAG/Cbp (phosphoprotein associated with GEMs/Csk-binding protein) phosphorylation recruiting Csk to LR to regulate Lck activity [34]. Consistent with this, strong TCR stimulation reduces constitutive PAG phosphorylation, thus favoring the dissociation of Csk, the main negative regulator of Lck activity. This is believed to induce TCR signal transduction.

Nrf2 activation alteration with aging

Aging is characterized by an increased level of oxidative stress. This also influences T cell activation. The mechanism of redox-dependent regulation of signaling pathways implies that receptor-mediated generation of reactive oxygen species (ROS) influences the balance between protein tyrosine kinase and phosphatase activities. Protein tyrosine kinases and phosphatases are redox-sensitive targets with an oxidation-sensitive active site cystein [37]. During receptor stimulation the transient increase of ROS may inactivate SHP-1 and thus facilitate TCR signaling (Figure 2A). However, in a setting where there is a constant increase in ROS, as in aging, SHP-1 regulation is altered and a negative regulatory role is favored in both the resting state and under stimulation. Another important constituent of the activation process is the cap 'n' collar basic region leucine zipper family



member nuclear factor erythroid 2-related factor -2 (Nrf2), a transcription factor [37]. Nrf2 is widely expressed in different cells and tissues, and is activated by oxidative and electrophilic stimulating agents, including ROS. Nrf2 modulates antioxidant gene expression by interacting with antioxidant response element. Nrf2 is tethered to the suppressor protein Kelch-like ECH-associated protein 1 in the cytoplasm under conditions of low oxidative stress, and is degraded by the ubiquitin-proteasome pathway. Under oxidative stress, the complex

dissociates, and Nrf2 translocates to the nucleus and enhances the expression of Nrf2/antioxidant response element-associated antioxidant genes [38]. Our own preliminary results suggest that Nrf2 is decreased upon stimulation of T cells from young individuals, indicating an increase of oxidative stress, while in the older people no changes in the Nrf2 levels were detected. This suggests a constantly increased ROS level, which is probably compensated for by alternative mechanisms inhibiting the translocation of Nrf2 into the nucleus (Figure 2B). This results in altered antioxidant generation, as has been demonstrated in T cells by the decrease of reduced glutathione and increase of oxidized glutathione levels [39]. Thus, with aging, increased oxidative stress together with the changes of tyrosine kinase and phosphatase activities, and their altered localization in the LR, contribute to altered T cell signaling with aging.

Role of lipid rafts in T cell signaling

Studies in model lipid bilayers have established that they can exist in an ordered (l_o) gel-like phase and a disordered (l_d) phase. In the case of the cell plasma membrane, the presence of cholesterol that associates with sphingolipids and other lipids allows the formation of microdomains in an l_o-like fluid phase that floats within the rest of the lipid bilayer, which is in the l_d phase. The term "lipid rafts" (LR) has been coined to designate these (l_0) microdomains [40,41]. LR are dynamic structures that are generally thought to be small, submicroscopic regions depleted of unsaturated phospholipids but enriched in cholesterol, alkyl chain-saturated sphingolipids and some lipid-anchored proteins [42]. Of significance, studies with lymphocytes have revealed that LR allow the recruitment of signaling proteins [43,44]. The coalescence of LR, dubbed "lipid rafting", is essential for the formation of the immune synapse and signal transduction in T cells [45]. However, this view was challenged by data from new imaging techniques such that conceptualization of the nature and function of LR is evolving [46-48]. Thus, LR are now considered as actual structures occupying spaces in the membrane as fluctuating dynamic nanoscale assemblies of sphingolipids, cholesterol and proteins that can be stabilized into platforms important for signaling and membrane trafficking. They can be further clustered into a second state by specific lipid-lipid, protein-lipid and protein-protein interactions [48] to generate more stable, selective and functional platforms. The merger of specific nanoscale rafts into larger and more stable platforms represents the functionalization of specific rafts in membrane trafficking as well as in signal transduction. For these events to occur the protein and lipid composition needs to be strictly fine tuned.

TCR are activated in microclusters subsequently transported to the locus of APC contact by actin filaments, generating a so-called central supramolecular activation cluster in which many TCRs are already dephosphorylated and inactive [49]. These data questioned the role of immune synapses in the signal transduction process, but helped to establish different roles for their components. This also helped to reconceptualize the composition and role of LR. Since then, modern high resolution imaging revealed the existence of complexes with up to 7 to 20 TCRs, manifested as protein islands 70 to 140 nm in diameter [50]. Were the resting states of raft proteins associated with nanoscale assemblies of raft lipids (for example, sphingolipids and cholesterol), such variances in size would result from the ability of these dynamic structures to coalesce. Stabilization of fluctuating nanoscale structures could facilitate increased access to Lck to mediate TCR signaling [33,51]. In this setting, how proteins and lipids (mainly cholesterol) interact remains an important question. How does the lipid context around the TCRs change the inactive to the activated state?

Role of cholesterol

The role of LR in T cell signaling has been challenged recently [47,48] despite substantial evidence in favor of their involvement in this process [46]. Several studies concluded that LR are indeed involved in T cell signaling because of the effects of acute cholesterol depletion - for example, both aggregation of plasma membrane LR and an increase in their numbers in the plasma membrane were noted on partial cholesterol depletion. This preceded Src family kinase activation, leading to increased co-localization between signaling molecules and resulting in an early T cell signaling response accompanied by marked actin polymerization in the plasma membrane [52]. It was also found that the kinase inhibitor PP2 did not prevent LR aggregation, which is in line with microcluster formation preceding T cell signaling [53]. The quantity of cholesterol extracted from the membrane can have differential effects on signal transduction, emphasizing that the lipid-protein interaction is as important as the protein-protein interaction in the signaling process.

Cholesterol is known to increase the thickness of lipid bilayers, as well as their stiffness. Both of these parameters are very important for interaction with membrane proteins. A series of investigations in the late 1980s had already suggested that biochemical and biophysical alterations at the level of the cell plasma membrane could be associated with the altered immune response with aging in mice [54,55]. It was noted that there were modifications with aging in the composition of the lymphocyte cell plasma membrane lipids as well as its fluidity. Earlier data from our laboratories have shown that the levels of plasma membrane cholesterol, the major stabilizing component of LR, were increased two-fold in T lymphocytes of older individuals, resulting in a plasma membrane that was less fluid than in T cells from young individuals [26]. We hypothesized that these observations had bearing on the events of T cell activation with aging. We investigated whether there were alterations in the proximal events of T cell activation upstream of the signaling cascade or at the very early events of signaling. We had used HPLC, thin layer chromatography and other lipid detecting techniques to show that there were marked changes in plasma membrane composition (fatty acids, phospholipids) in T cells of middle-aged normolipemic subjects [56]. We put forward the hypothesis that these changes could be related to the decreased T cell functions associated with immunosenescence. We attempted to modulate the levels of plasma membrane cholesterol in T cells from older subjects in vitro to determine whether this manipulation re-established efficient T cell signaling/functions. Initial experiments using the cholesterol sequestrating compound methyl β-cyclodextrin (MBCD) did not yield results expected on the basis of the hypothesis mentioned above. It was observed that, besides decreasing plasma membrane cholesterol, MBCD had an independent signal-disrupting activity. We therefore also used inhibitors of cholesterol synthesis (lovastatin) in Jurkat T cells. In this case, significantly reduced cell growth and induced apoptosis at high concentrations were observed, in agreement with the fact that only a few studies have demonstrated the immunomodulatory role of statins in human T cells [57]. Reciprocally, we attempted to "age" T cells of young subjects by increasing the cholesterol content to the levels found in T cells of older subjects. This led to a decreased proliferative response as well as IL-2 secretion, suggesting that homeostatic concentrations of plasma membrane cholesterol are important for T cell activation (Figure 3). An attractive approach in vivo might therefore be to use high-density lipoprotein (HDL) as a physiologic means to extract cholesterol by the so-called reverse transport cholesterol mechanism. HDL particles or ApoA-I interact with cells to promote the efflux of cholesterol via binding to specific transporters of the ATP-binding cassette (ABC) gene family [58].



ABCG1 induces cholesterol efflux to HDL particles [59]. HDL preferentially interacts with LR and depletes cholesterol from these microdomains [60]. By this extraction effect, HDL may modulate the signaling and the functions of T cells. More work is needed to determine the exact role of cholesterol and HDL in the altered activation of T cells with aging.

There are no well documented molecular mechanisms to account for increased plasma membrane cholesterol content in T cells of normolipemic older subjects. However, a dysregulation of cholesterol uptake involving the low-density lipoprotein receptor, an increase in the synthesis of cholesterol resulting from upregulation of HMG-CoA reductase activity, an alteration in the activity of the ACAT enzyme or a deficiency in HDLdependent reverse cholesterol transport are all possible causes. Further studies of cholesterol metabolism in T cells and the modulation of the cholesterol content of the plasma membrane are needed to shed light on the alterations and the functional changes of LR and the early events of T cell activation with aging.

Alterations in lipid raft functions with aging

LR move across the plasma membrane lipid bilayer as discrete complexes to assemble the signalosome [61] prior or concomitantly to the formation of the immune synapse. LR clustering may depend on GM1 (monosialotetrahexosylganglioside), GM3 and flotillin-1 [62] as well as reorganization of the cytoskeleton. Whereas some of the signaling components are constitutive parts of LR (Lck, Cbp/PAG), others are recruited during activation (TCR, CD28, IL-2 receptor, LAT, PI-3 K). We were the first group to report a study of LR alterations in older humans with aging [63]. We have shown that LR are distributed homogeneously in T cells of young subjects, whereas we observed a different LR pattern and coalescence in the quiescent state in T cells of older subjects [26]. These observations may reflect, to some extent, low-grade activation with aging as part of the "Inflamm-Aging" model [64]. We have also reported an alteration in the function of LR with aging. Using confocal microscopy, we have shown that LR coalescence is altered with aging in T cells activated by a combination of anti-TCR and anti-CD28 antibodies. LR poorly coalesced in CD4⁺ T cells of older subjects [22] although the alterations were less pronounced in the case of CD8⁺ T cells. Furthermore, we have reported alterations in the recruitment and activation of Lck and LAT into LR of T cells from older humans [26]. One important finding was that CD28 and the IL-2 receptor were weakly recruited to LR in CD4⁺ T cells of older subjects. In contrast, these proteins were already located in LR in CD8⁺ T cells from older subjects prior to stimulation. These observations suggested that the assembly of the signaling machinery

in $CD4^+$ T cells relies largely on LR, whereas a preassembly of the signalosome may be present in $CD8^+$ T lymphocytes, as proposed by us [22] and by others [65,66]. Currently we do not know why this alteration of the plasma membrane occurs with aging. An intrinsic alteration of cholesterol metabolism in older subjects could contribute to these lipid changes [67]. Therefore, an alteration in the plasma membrane composition of T cells with aging seems to play a fundamental role in their decreased functions as observed in immunosenescence. Moreover, an alteration of the cytoskeleton in T cells with aging can further contribute to altered signaling *via* altered LR functions.

Conclusions

Increasing numbers of older individuals not only in industrialized nations represents major social and medical challenges. Concomitantly, the incidence of agerelated diseases, many of which are influenced by the dysregulation of the immune system, is increasing. There is a large corpus of experimental data suggesting that the immune response, mainly the T cell response, is dysregulated in aging. The causes of this dysregulation remain largely unknown, although evidence for alterations in T cell receptor and CD28 co-receptor signaling is widely accepted. One unifying cause for these alterations could involve the initial events of signal transduction and the role of the plasma membrane. The novel concept that the plasma membrane is composed of privileged signal transduction microdomains (LR) that create a functional dynamic environment to assemble the signalosome and the constitution of the immune synapse provides a working framework to address the molecular mechanisms of immunosenescence. Changes in the lipid composition of the plasma membrane, especially in LR, can be envisaged as a key event that adversely affects T lymphocyte signaling in older subjects. These functional changes may have serious clinical consequences such as increased infections, cancers, autoimmune diseases and thus poor quality of life. According to one estimate, more than 50 pathologies have been associated with dysfunctional LR [68]. The inter-individual susceptibility to the diseases of aging may be the cause or the consequence of the differential changes in the immune response (which is associated with T cell pool composition). It is of fundamental importance to understand the role of LR to be able to intervene more precisely during the evolution of diseases affecting older people. New imaging techniques should be applied to the study of signaling events in T cells of healthy older subjects [46,47]. These studies could lead to the elaboration of new intervention tools in the management of the physiological as well as the pathological process of aging.

Data from our laboratories and others have provided a basis to pursue studies to understand the molecular mechanisms of T cell signaling deficiencies associated with aging, the involvement of LR, the formation of the immune synapse and the assembly of the signaling machinery. Moreover, investigating T lymphocyte cholesterol metabolism will help to understand the increase of cholesterol content in the membrane. The role of free radicals should also be assessed in connection with anti-oxidant defense. Thus, a better understanding of immunosenescence by intensive research and the development of new methods and strategies to intervene in its evolution are essential for improving the quality of life of an increasingly larger older population.

Abbreviations

ABC: ATP-binding cassette; APC: antigen-presenting cell; Cbp: Csk-binding protein; Csk: C-terminal Src kinase; HDL: high-density lipoprotein; HPLC: high performance liquid chromatography; IL: interleukin; LR: lipid rafts; MBCD: methyl β-cyclodextrin; Nrf2: nuclear factor erythroid 2-related factor-2; PAG: phosphoprotein associated with GEMs; ROS: reactive oxygen species; SHP-1: *Src* homology 2 domain-containing protein tyrosine-phosphatase 1; TCR: T cell receptor; TNF: tumor necrosis factor; ZAP70: Zeta-chain-associated protein kinase 70.

Competing interests

The authors declare that they have no competing interests.

Authors' contributions

All authors contributed to writing the manuscript. All authors read and approved the final manuscript.

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References

- 1. Agarwal S, Busse PJ: Innate and adaptive immunosenescence. Ann Allergy Asthma Immunol 2010, 104:183–190.
- Fulop T, Larbi A, Wikby A, Mocchegiani E, Hirokawa K, Pawelec G: Dysregulation of T-cell function in the elderly: scientific basis and clinical implications. *Drugs Aging* 2005, 22:589–603.
- Schoenborn JR, Tan YX, Zhang C, Shokat KM, Weiss A: Feedback circuits monitor and adjust basal Lck-dependent events in T cell receptor signaling. Sci Signal 2011, 4(190):ra59.
- Larbi A, Pawelec G, Wong SC, Goldeck D, Tai JJ, Fulop T: Impact of age on T cell signaling: a general defect or specific alterations? *Ageing Res Rev* 2011, 10:370–378.

- Alarcón B, Mestre D, Martínez-Martín N: The immunological synapse: a cause or consequence of T-cell receptor triggering? *Immunology* 2011, 133:420–425.
- 6. Reichardt P, Dornbach B, Gunzer M: APC, T cells, and the immune synapse. *Curr Top Microbiol Immunol* 2010, 340:229–249.
- Rodríguez-Fernández JL, Riol-Blanco L, Delgado-Martín C: What is an immunological synapse? *Microbes Infect* 2010, 12:438–445.
- 8. Manes S, Viola A: Lipid rafts in lymphocyte activation and migration. Mol Membr Biol 2006, 23:59–69.
- Wick G, Jansen-Dürr P, Berger P, Blasko I, Grubeck-Loebenstein B: Diseases of aging. Vaccine 2000, 18:1567–1583.
- 10. Meyer KC: Aging. Proc Am Thorac Soc 2005, 2:433-439.
- Pawelec G: Immunosenescence and human longevity. Biogerontol 2003, 4:167–170.
- Ligthart GH: The SENIEUR protocol after 16 years: the next step is to study the interaction of ageing and disease. *Mech Ageing Dev* 2001, 122:136–140.
- 13. Fulop T, Larbi A, Kotb R, de Angelis F, Pawelec G: Aging, immunity, and cancer. *Discov Med* 2011, 11:537–550.
- 14. Vasto S, Malavolta M, Pawelec G: Age and immunity. Immun Ageing 2006, 3:2.
- Solana R, Tarazona R, Gayoso I, Lesur O, Dupuis G, Fulop T: Innate immunosenescence: effect of aging on cells and receptors of the innate immune system in humans. *Semin Immunol* 2012, [Epub ahead of print].
- Smith-Garvin JE, Koretzky GA, Jordan MS: T cell activation. Annu Rev Immunol 2009, 27:591–619.
- Nel AE, Slaughter N: T-cell activation through the antigen receptor. Part 2: role of signaling cascades in T-cell differentiation, anergy, immune senescence, and development of immunotherapy. J Allergy Clin Immunol 2002, 109:901–915.
- Riley JL, June CH: The CD28 family: a T-cell rheostat for therapeutic control of T-cell activation. *Blood* 2005, 105:13–21.
- Bjørgo E, Taskén K: Novel mechanism of signaling by CD28. Immunol Lett 2010, 129:1–6.
- 20. Tavano R, Gri G, Molon B, Marinari B, Rudd CE, Tuosto L, Viola A: CD28 and lipid rafts coordinate recruitment of Lck to the immunological synapse of human T lymphocytes. *J Immunol* 2004, **173**:5392–5397.
- Wang D, Matsumoto R, You Y, Che T, Lin XY, Gaffen SL, Lin X: CD3/CD28 costimulation-induced NF-kappaB activation is mediated by recruitment of protein kinase C-theta, Bcl10, and IkappaB kinase beta to the immunological synapse through CARMA1. *Mol Cell Biol* 2004, 24:164–171.
- Larbi A, Dupuis G, Khalil A, Douziech N, Fortin C, Fulop T Jr: Differential role of lipid rafts in the functions of CD4+ and CD8+ human T lymphocytes with aging. *Cell Signal* 2006, 18:1017–1030.
- 23. Sadighi Akha AA, Miller RA: Signal transduction in the aging immune system. *Curr Opin Immunol* 2005, **17**:486–491.
- Garcia GG, Miller RA: Age-related defects in CD4+ T cell activation reversed by glycoprotein endopeptidase. Eur J Immunol 2003, 33:3464–3472.
- Vallejo AN, Brandes JC, Weyand CM, Goronzy JJ: Modulation of CD28 expression: distinct regulatory pathways during activation and replicative senescence. J Immunol 1999, 162:6572–6579.
- Larbi A, Douziech N, Dupuis G, Khalil A, Pelletier H, Guerard KP, Fülöp T: Age-associated alterations in the recruitment of signal-transduction proteins to lipid rafts in human T lymphocytes. J Leukoc Biol 2004, 75:373–381.
- Ponnappan U, Trebilcock GU, Zheng MZ: Studies into the effect of tyrosine phosphatase inhibitor phenylarsine oxide on NFkappaB activation in T lymphocytes during aging: evidence for altered lkappaBalpha phosphorylation and degradation. *Exp Gerontol* 1999, 34:95–107.
- Rink L, Cakman I, Kirchner H: Altered cytokine production in the elderly. Mech Ageing Dev 1998, 102:199–209.
- Acuto O, Di Bartolo V, Michel F: Tailoring T-cell receptor signals by proximal negative feedback mechanisms. *Nature Rev Immunol* 2008, 8:699–712.
- Iwashima M, Irving BA, van Oers NS, Chan AC, Weiss A: Sequential interactions of the TCR with two distinct cytoplasmic tyrosine kinases. *Science* 1994, 263:1136–1139.
- Yamaguchi H, Hendrickson WA: Structural basis for activation of human lymphocyte kinase Lck upon tyrosine phosphorylation. *Nature* 1996, 384:484–489.

- Burns CM, Sakaguchi K, Appella E, Ashwell JD: CD45 regulation of tyrosine phosphorylation and enzyme activity of src family kinases. *J Biol Chem* 1994, 269:13594–13600.
- Nika K, Soldani C, Salek M, Paster W, Gray A, Etzensperger R, Fugger L, Polzella P, Cerundolo V, Dushek O, Höfer T, Viola A, Acuto O: Constitutively active Lck kinase in T cells drives antigen receptor signal transduction. *Immunity* 2010, 32:766–777.
- Yasuda K, Nagafuku M, Shima T, Okada M, Yagi T, Yamada T, Minaki Y, Kato A, Tani-Ichi S, Hamaoka T, Kosugi A: Cutting edge: Fyn is essential for tyrosine phosphorylation of Csk-binding protein/phosphoprotein associated with glycolipid-enriched microdomains in lipid rafts in resting T cells. J Immunol 2002, 169:2813–2817.
- 35. Lorenz U: SHP-1 and SHP-2 in T cells: two phosphatases functioning at many levels. *Immunol Rev* 2009, **228**:342–359.
- Barrett DM, Black SM, Todor H, Schmidt-Ullrich RK, Dawson KS, Mikkelsen RB: Inhibition of protein-tyrosine phosphatases by mild oxidative stresses is dependent on S-nitrosylation. J Biol Chem 2005, 280:14453–14461.
- Li H, Wang F, Zhang L, Cao Y, Liu W, Hao J, Liu Q, Duan H: Modulation of Nrf2 expression alters high glucose-induced oxidative stress and antioxidant gene expression in mouse mesangial cells. *Cell Signal* 2011, 23:1625–1632.
- McMahon M, Thomas N, Itoh K, Yamamoto M, Hayes JD: Dimerization of substrate adaptors can facilitate cullin-mediated ubiquitylation of proteins by a "tethering" mechanism: a two-site interaction model for the Nrf2-Keap1 complex. J Biol Chem 2006, 281:24756–24768.
- Larbi A, Cabreiro F, Zelba H, Marthandan S, Combet E, Friguet B, Petropoulos I, Barnett Y, Pawelec G: Reduced oxygen tension results in reduced human T cell proliferation and increased intracellular oxidative damage and susceptibility to apoptosis upon activation. *Free Radic Biol Med* 2010, 48:26–34.
- Simons K, Ikonen E: Functional rafts in cell membranes. Nature 1997, 387:569–572.
- Brown DA, London E: Functions of lipid rafts in biological membranes. Annu Rev Cell Dev Biol 1998, 14:111–136.
- 42. Brown DA: Lipid rafts, detergent-resistant membranes, and raft targeting signals. *Physiology (Bethesda)* 2006, **21**:430–439.
- Pizzo P, Viola A: Lipid rafts in lymphocyte activation. Microbes Infect 2004, 6:686–692.
- Kabouridis PS: Lipid rafts in T cell receptor signalling. Mol Membr Biol 2006, 23:49–57.
- Taner SB, Onfelt B, Pirinen NJ, McCann FE, Magee AI, Davis DM: Control of immune responses by trafficking cell surface proteins, vesicles and lipid rafts to and from the immunological synapse. *Traffic* 2004, 5:651–661.
- 46. Tolar P: Inside the microcluster: antigen receptor signalling viewed with molecular imaging tools. *Immunology* 2011, **133**:271–277.
- 47. Simons K, Gerl MJ: Revitalizing membrane rafts: new tools and insights. Nature Rev Mol Cell Biol 2010, 11:688–699.
- Simons K, Sampaio JL: Membrane organization and lipid rafts. Cold Spring Harb Perspect Biol 2011, 3:a004697.
- Kaizuka Y, Douglass AD, Varma R, Dustin ML, Vale RD: Mechanisms for segregating T cell receptor and adhesion molecules during immunological synapse formation in Jurkat T cells. Proc Natl Acad Sci U S A 2007, 104:20296–20301.
- Lillemeier BF, Mörtelmaier MA, Forstner MB, Huppa JB, Groves JT, Davis MM: TCR and Lat are expressed on separate protein islands on T cell membranes and concatenate during activation. *Nat Immunol* 2010, 11:90–96.
- Irles C, Arias-Martinez J, Guzmán-Bárcenas J, Ortega A: Plasma membrane subdomain partitioning of Lck in primary human T lymphocytes. Can J Physiol Pharmacol 2010, 88:487–496.
- Mahammad S, Dinic J, Adler J, Parmryd I: Limited cholesterol depletion causes aggregation of plasma membrane lipid rafts inducing T cell activation. *Biochim Biophys Acta* 2010, 1801:625–634.
- Billadeau DD, Burkhardt JK: Regulation of cytoskeletal dynamics at the immune synapse: new stars join the actin troupe. *Traffic* 2006, 7:1451–1460.
- 54. Maczek C, Bock G, Jurgens G, Schonitzer D, Dietrich H, Wick G: Environmental influence on age-related changes of human lymphocyte

membrane viscosity using severe combined immunodeficiency mice as an *in vivo* model. *Exp Gerontol* 1998, **33**:485–498.

- Huber LA, Xu QB, Jürgens G, Böck G, Bühler E, Gey KF, Schönitzer D, Traill KN, Wick G: Correlation of lymphocyte lipid composition membrane microviscosity and mitogen response in the aged. *Eur J Immunol* 1991, 21:2761–2765.
- Seres I, Freyss-Beguin M, Mohacsi A, Kozlovsky B, Simon J, Devynck MA, Fulop T Jr: Alteration of lymphocyte membrane phospholipids and intracellular free calcium concentrations in hyperlipidemic subjects. *Atherosclerosis* 1996, **121:**175–183.
- Fulop T, Dupuis G, Fortin C, Douziech N, Larbi A: T cell response in aging: influence of cellular cholesterol modulation. *Adv Exp Med Biol* 2006, 584:157–169.
- Ansell BJ, Watson KE, Fogelman AM, Navab M, Fonarow GC: High-density lipoprotein function recent advances. J Am Coll Cardiol 2005, 46:1792–1798.
- Wang N, Lan D, Chen W, Matsuura F, Tall A: ATP-binding cassette transporters G1 and G4 mediate cellular cholesterol efflux to high-density lipoproteins. Proc Natl Acad Sci U S A 2004, 101:9774–9779.
- Yu BL, Wang SH, Peng DQ, Zhao SP: HDL and immunomodulation: an emerging role of HDL against atherosclerosis. *Immunol Cell Biol* 2010, 88:285–290.
- 61. Cemerski S, Shaw A: Immune synapses in T-cell activation. Curr Opin Immunol 2006, 18:298–304.
- Douglass AD, Vale RD: Single-molecule microscopy reveals plasma membrane microdomains created by protein-protein networks that exclude or trap signaling molecules in T cells. *Cell* 2005, 121:937–950.
- Fülöp T, Douziech N, Goulet AC, Desgeorges S, Linteau A, Lacombe G, Dupuis G: Cyclodextrin modulation of T lymphocyte signal transduction with aging. *Mech Age Dev* 2001, 122:1413–1430.
- 64. Larbi A, Dupuis G, Douziech N, Khalil A, Fulop T: Low-grade inflammation with aging has consequences for T-lymphocyte signaling. *Ann NY Acad Sci* 2004, **1030**:125–133.
- Kovacs B, Maus MV, Riley JL, Derimanov GS, Koretzky GA, June CH, Finkel TH: Human CD8+ T cells do not require the polarization of lipid rafts for activation and proliferation. *Proc Natl Acad Sci USA* 2002, 99:15006–15011.
- de Mello Coelho V, Nguyen D, Giri B, Bunbury A, Schaffer E, Taub DD: Quantitative differences in lipid raft components between murine CD4+ and CD8+ T cells. *BMC Immunol* 2004, 5:2.
- Simons K, Ikonen E: How cells handle cholesterol. Science 2000, 290:1721–1726.
- Michel V, Bakovic M: Lipid rafts in health and disease. Biol Cell 2007, 99:129–140.

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